illumina®

Infinium Multi-Use LCG Assay
Lab Tracking Form

1 Make and Read Quant (Optional/LIMS)

Date/Time: _____

Operator: _____ Robot: _____

WG#-DNA Plate: QNT Plate:

(1) _____ _____

(2) _____ _____

(3) _____ _____

Standard DNA Plate: Standard QNT Plate:

_____ _____

2 Make AMP4

Date/Time: _____

Operator: _____ Robot: _____

Plate Positions on Robot Bed: _____

Batch #: _____

Number of Samples: _____

Hyb oven (37°C, 20-24 h): Start _____ Stop _____

WG#-DNA Plate: _____

0.1N NaOH Reagent: _ _____

AMP4 Plate: _____

RPM Reagent: (Col. 1) _____

(Col. 5) _____

(Col. 9) _____

AMM Reagent: (Col. 1) _____

(Col. 5) _____

(Col. 9) _____

Column 1 Well: Sample	Column 5 Well: Sample	Column 9 Well: Sample
A01: _____	A05: _____	A09: _____
B01: _____	B05: _____	B09: _____
C01: _____	C05: _____	C09: _____
D01: _____	D05: _____	D09: _____
E01: _____	E05: _____	E09: _____
F01: _____	F05: _____	F09: _____
G01: _____	G05: _____	G09: _____
H01: _____	H05: _____	H09: _____

3 Fragment AMP4

Date/Time: _____

Operator: _____ Robot: _____

Plate Positions on Robot Bed: _____

☒ Vortex at 1600 rpm

Heat block (37°C, 1 h): Start _____ Stop _____

FRG Reagent: (Col. 1) _____

(Col. 5) _____

(Col. 9) _____

Project: _____ Batch: _____ AMP4 Plate: _____ Image Date: _____

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4 Precip AMP4

Date/Time: _____

Operator: _____ Robot: _____

Plate Positions on Robot Bed: _____

☐ Vortex at 1600 rpm

Heat block (37°C, 5m): _____

Incubate (4°C, 30m): Start: _____ Stop: _____

Air dry (22°C, 1 h): Start: _____ Stop: _____

2-propanol Lot #: _____

2-propanol Date Opened: _____

PA1 Reagent: (Col. 1) _____

(Col. 5) _____

(Col. 9) _____

5 Resuspend AMP4

Date/Time: _____

Operator: _____ Robot: _____

Plate Positions on Robot Bed: _____

Hyb oven (48°C, 1 h): Start _____ Stop _____

RA1 Reagent: _____

First Hybridization

6 Hyb Multi-Use BeadChip

Date/Time: _____
Operator: _____ Robot: _____
☐ Vortex at 1800 rpm
Heat block (95°C, 20m): Start: _____ Stop: _____
☐ Centrifuge AMP4 plate to 280 xg
Hyb oven (48°C, 16-24 h): Start _____ Stop _____

PB2 Reagent: _____
_____
_____

Refer to page 4 for BeadChip loading instructions. Enter the BeadChip barcodes in the spaces provided.

7 Wash BeadChip

Date/Time: _____
Operator: _____ Robot: _____

PB1 Reagent: _____
_____
_____

8 XStain LCG BeadChip

Date/Time: _____
Operator: _____ Robot: _____
RA1 Reagent: _____
LX1 Reagent: (1-8) _____
(9-16) _____
(17-24) _____
LX2 Reagent: (1-8) _____
(9-16) _____
(17-24) _____
EML Reagent: (1-8) _____
(9-16) _____
(17-24) _____
95% formamide/1 mM EDTA Reagent: _____

XC3 Reagent: _____
SML Reagent: (1-8) _____
(9-16) _____
(17-24) _____
SML Temperature: _____
ATM Reagent: (1-8) _____
(9-16) _____
(17-24) _____
PB1 Reagent: _____
XC4 Reagent: _____

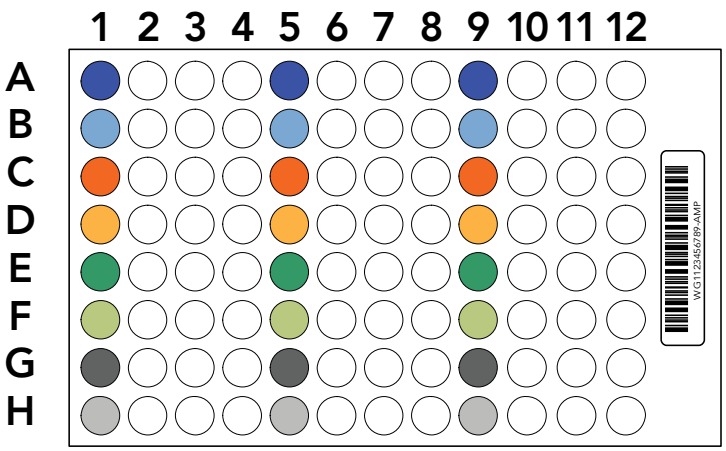
Record the chamber rack position for each BeadChip on page 7.

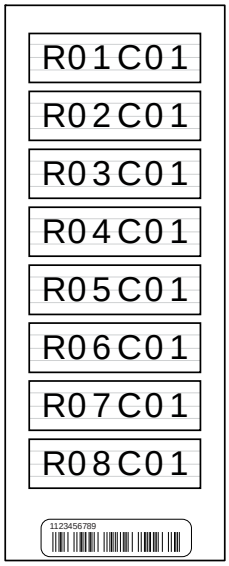
9 Image BeadChip

Record the Scanner ID and the image date for each BeadChip on the appropriate BeadChip tracking page.

Track BeadChips (First Hybridization)

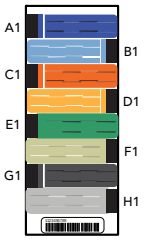
AMP Plate





Sample Section Naming Diagram

BC2 #1

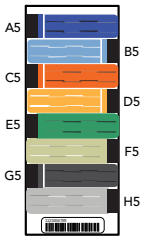


Barcode: _____

Scanner ID: _____

Image Date: _____

BC2 #2

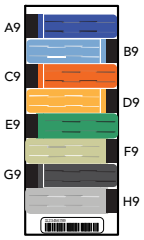


Barcode: _____

Scanner ID: _____

Image Date: _____

BC2 #3



Barcode: _____

Scanner ID: _____

Image Date: _____

Second Hybridization

6 Hyb Multi-Use BeadChip

Date/Time: _____
Operator: _____ Robot: _____
☐ Vortex at 1800 rpm
Heat block (95°C, 20m): Start: _____ Stop: _____
☐ Centrifuge AMP4 plate to 280 xg
Hyb oven (48°C, 16-24 h): Start _____ Stop _____

PB2 Reagent: _____
_____
_____
AMPx Plate: _____

Refer to page 6 for BeadChip loading instructions. Enter the BeadChip barcodes in the spaces provided.

7 Wash BeadChip

Date/Time: _____
Operator: _____ Robot: _____

PB1 Reagent: _____
_____
_____

8 XStain LCG BeadChip

Date/Time: _____
Operator: _____ Robot: _____
RA1 Reagent: _____
LX1 Reagent: (1-8) _____
(9-16) _____
(17-24) _____
LX2 Reagent: (1-8) _____
(9-16) _____
(17-24) _____
EML Reagent: (1-8) _____
(9-16) _____
(17-24) _____
95% formamide/1 mM EDTA Reagent: _____

XC3 Reagent: _____
SML Reagent: (1-8) _____
(9-16) _____
(17-24) _____
SML temperature: _____
ATM Reagent: (1-8) _____
(9-16) _____
(17-24) _____
PB1 Reagent: _____
XC4 Reagent: _____

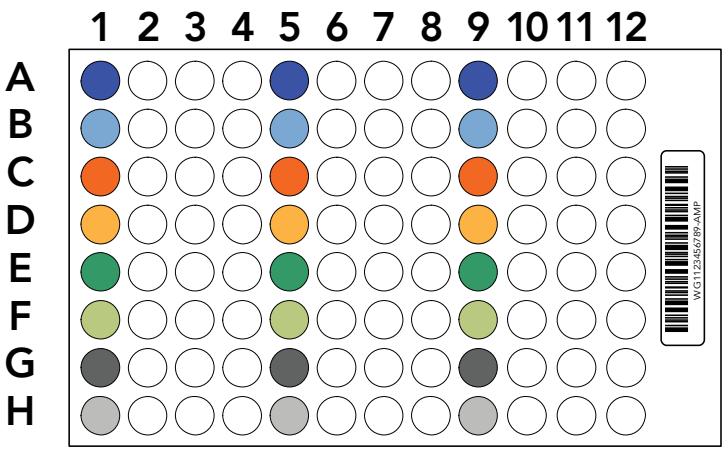
Record the chamber rack position for each BeadChip on page 7.

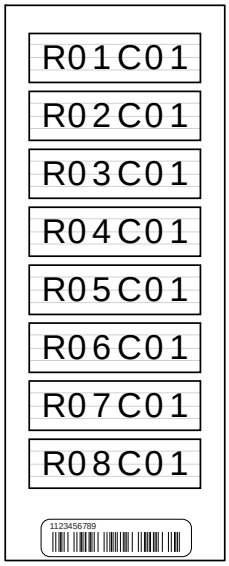
9 Image BeadChip

Record the Scanner ID and the image date for each BeadChip on the appropriate BeadChip tracking page.

Track BeadChips (Second Hybridization)

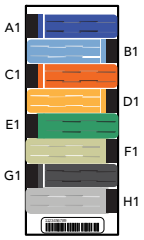
AMP Plate





Sample Section Naming Diagram

BC2 #1

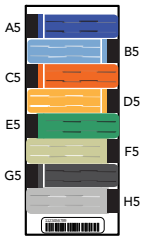


Barcode: _____

Scanner ID: _____

Image Date: _____

BC2 #2

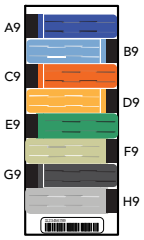


Barcode: _____

Scanner ID: _____

Image Date: _____

BC2 #3



Barcode: _____

Scanner ID: _____

Image Date: _____

Third Hybridization

6 Hyb Multi-Use BeadChip

Date/Time: _____
Operator: _____ Robot: _____
☐ Vortex at 1800 rpm
Heat block (95°C, 20m): Start: _____ Stop: _____
☐ Centrifuge AMP4 plate to 280 xg
Hyb oven (48°C, 16-24 h): Start _____ Stop _____

PB2 Reagent: _____
_____
_____
AMPx Plate: _____

Refer to page 4 for BeadChip loading instructions. Enter the BeadChip barcodes in the spaces provided.

7 Wash BeadChip

Date/Time: _____
Operator: _____ Robot: _____

PB1 Reagent: _____
_____
_____

8 XStain LCG BeadChip

Date/Time: _____
Operator: _____ Robot: _____
RA1 Reagent: _____
LX1 Reagent: (1-8) _____
(9-16) _____
(17-24) _____
LX2 Reagent: (1-8) _____
(9-16) _____
(17-24) _____
EML Reagent: (1-8) _____
(9-16) _____
(17-24) _____
95% formamide/1 mM EDTA Reagent: _____

XC3 Reagent: _____
SML Reagent: (1-8) _____
(9-16) _____
(17-24) _____
SML Temperature: _____
ATM Reagent: (1-8) _____
(9-16) _____
(17-24) _____
PB1 Reagent: _____
XC4 Reagent: _____

Record the chamber rack position for each BeadChip on page 7.

9 Image BeadChip

Record the Scanner ID and the image date for each BeadChip on the appropriate BeadChip tracking page.

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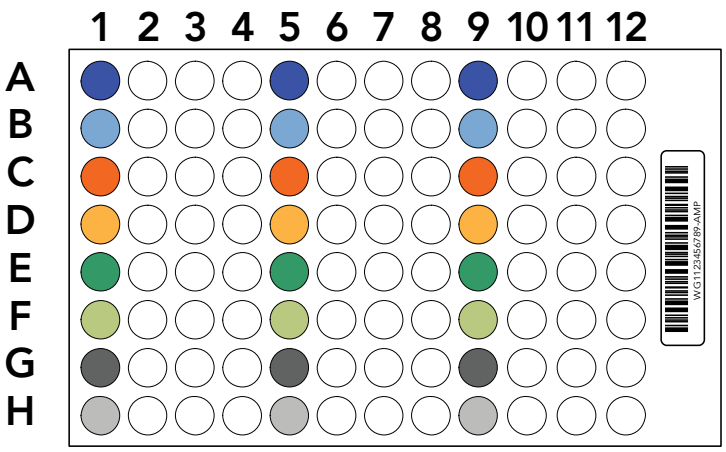
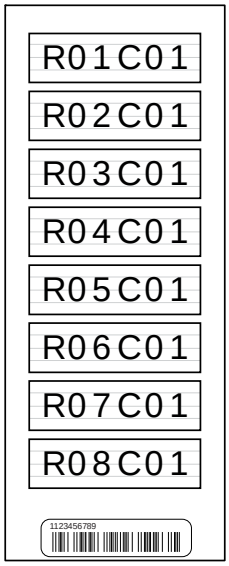
Track BeadChips (Third Hybridization)

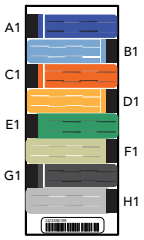
AMP Plate





Sample Section Naming Diagram

BC2 #1

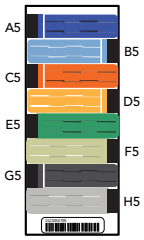


Barcode: _____

Scanner ID: _____

Image Date: _____

BC2 #2

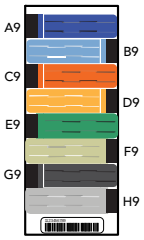


Barcode: _____

Scanner ID: _____

Image Date: _____

BC2 #3



Barcode: _____

Scanner ID: _____

Image Date: _____

Chamber Rack Position Chart

Use this chart to enter BeadChip IDs in the appropriate chamber rack position during the XStain LCG BeadChip step.

Row 1	Row 2	Row 3
1 _____	9 _____	17 _____
2 _____	10 _____	18 _____
3 _____	11 _____	19 _____
4 _____	12 _____	20 _____
5 _____	13 _____	21 _____
6 _____	14 _____	22 _____
7 _____	15 _____	23 _____
8 _____	16 _____	24 _____